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Hydrolysis of Enkephalin by Cultured
Human Endothelial Cells and by Purified Peptidyl Dipeptidase

Prepared by

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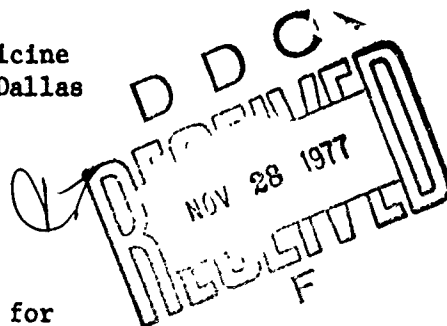
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The biological activities of enkephalins are of great interest to numerous investigators since they have an affinity for opiate receptors and may mediate perception of pain ¹. Studies of enkephalins have been hampered, however, because these endogenous pentapeptides are rapidly degraded by blood and tissues. For example, radiolabeled enkephalin is hydrolyzed in less than one minute when it is injected intraventricularly in rats ².

Peptides transported in blood are continuously in contact with the vascular endothelium. These cells are more than just a diffusion barrier between blood and tissues since they contain enzymes that metabolize various peptides. Enzymes in vascular endothelial cells grown in culture cleave peptides such as bradykinin, angiotensin I and II ³ and substance P ⁴. Two of the enzymes in cultured human endothelial cells were identified as an aminopeptidase and a peptidyl dipeptidase ³. These two enzymes have important functions in the control of both hypotensive and hypertensive peptides. The aminopeptidase may hydrolyze angiotensin I or II by cleaving the N-terminal aspartic acid. The peptidyl dipeptidase cleaves a dipeptide from the C-terminal end of bradykinin and angiotensin I. By this action it converts angiotensin I to II and inactivates bradykinin ^{5,6}. In addition to endothelial cells, the brush border of the kidney ⁶ and various parts of the central nervous system contain peptidyl dipeptidase activity ^{7,8}. Because the substrate specificity of this enzyme (E.C. 3.4.15.1; kininase II or angiotensin I converting enzyme) indicated ⁶ that enkephalins may also be cleaved, we investigated the action of purified enzyme on synthetic leucine- and methionine-enkephalins. We determined the peptide bond hydrolyzed and also characterized the mode of inactivation of enkephalins by cultured human endothelial cells.

Leu⁵-enkephalin (Tyr¹-Gly²-Gly³-Phe⁴-Leu⁵; Leu-Enk) and Met⁵-enkephalin (Met-

enk) were obtained from commercial sources. Endothelial cells from veins of human umbilical cords were grown in monolayer cell cultures as described ³. Peptidyl dipeptidase was purified to homogeneity by the method of Oshima et al. ⁹. Leucine aminopeptidase (E.C. 3.4.1.1) was purchased from Boehringer GmbH. Leu-enk or Met-enk (0.25 μmol) was incubated with 1.2 μg peptidyl dipeptidase or 0.12 μg aminopeptidase at 37° in 0.06 M NH_4HCO_3 buffer, pH 7.4. The molar ratio of peptidyl dipeptidase to substrate was 1 to 30,000. Cultured human endothelial cells were removed from monolayers by gentle scraping with a rubber spatula. These cells were then suspended and used in a concentration of one million cells per ml. The products of hydrolysis of enkephalins were determined by thin layer chromatography (t.l.c.).

The rate of cleavage of Met-enk by peptidyl dipeptidase was determined in a programmed amino acid analyzer by separating Tyr-Gly-Gly and Phe-Met on a 5.5 cm column in 0.2 N citrate buffer, pH 4.25. The column was calibrated with the synthetic tripeptide and dipeptide. Substrate (0.2 $\mu\text{mol/ml}$) and enzyme (2.4 $\mu\text{g/ml}$) were incubated at 37° and pH 7.4 in 0.1 M Tris containing 0.1 M NaCl. In control studies the cleavage of enkephalins was completely inhibited by one of the specific inhibitors of the enzyme: either by 10^{-4}M of the non-peptide SQ 20881 ⁶ or by 10^{-6}M of SQ 14225 (2-D-methyl-3-mercapto-L-proline) ¹⁰.

Both Leu-enk and Met-enk were cleaved by peptidyl dipeptidase at the Gly³-Phe⁴ bond to Tyr¹-Gly²-Gly³ and to Phe⁴-Leu⁵ or Phe⁴-Met⁵. Figure 1 shows the separation of the peptides in t.l.c. in two different systems. Fig. 1

The rate of hydrolysis of Met-enk was measured with an amino acid analyzer (Fig. 2). The enzyme and the substrate were incubated in a buffer more favor- Fig. 2

able for enzyme action than the one used for t.l.c. Instead of the NH_4HCO_3 buffer, the incubation medium contained 0.1 M NaCl in Tris buffer, since the enzyme cleaves most substrates faster in presence of Cl^- ⁵. Peptidyl dipeptidase cleaved Met-enk at a rate of 4.2 $\mu\text{mol}/\text{min}$ per mg protein and not at all in presence of SQ 20881. The same enzyme preparation cleaved the specific substrate Bz-Gly-Gly-Gly (10^{-3}M) at a rate of 7.4 $\mu\text{mol}/\text{min}$ when determined in the u.v. spectrophotometer ⁵.

Because cultured endothelial cells contain both aminopeptidase- and peptidyl dipeptidase-type enzymes we utilized t.l.c. to determine whether the N or C-terminal end of enkephalin is hydrolyzed first by enzymes of these cells.

Figure 3a shows the separation of products of hydrolysis of Leu-enk by endothelial cells. The $\text{Tyr}^1\text{-Gly}^2$ bond was cleaved first by the endothelial aminopeptidase. The enzyme was inhibited by the sequestering agent o-phenanthroline (10^{-3}M). In control studies a highly purified commercial leucine aminopeptidase (E.C. 3.4.11.1) was used (Fig. 3b) which also cleaved the pentapeptide to Tyr^1 and to tetrapeptide. We could detect the release of Phe-Leu by the action of endothelial peptidyl dipeptidase only after an incubation of 60 min or longer. Fig. 3.

These experiments have shown that purified peptidyl dipeptidase cleaved enkephalins to tri- and dipeptide. The unprotected tripeptide liberated may be resistant to further degradation by the enzyme because of the free $\alpha\text{-NH}_2$ group.

Peptidyl dipeptidase is concentrated in various parts of the body, for example, in vascular endothelium of the lung and other organs, in kidney, in testicles ⁶, and pituitary gland ⁷. In the central nervous system it is present in structures such as choroid plexus ¹¹, caudate nucleus ⁸ or striatum ⁷.

The rate of cleavage of Met-enk by peptidyl dipeptidase is quite substantial.

Under our conditions the rate was more than half of that of hydrolysis of Bz-Gly-Gly-Gly, the specific substrate of this enzyme,^{5,6} although the concentration of enkephalin was 1/5 of that of the optically active tripeptide. Since only peptides with free C-terminal end are cleaved by peptidyl dipeptidase⁶ resistance to the action of the enzyme may be one of the reasons for the markedly enhanced in vivo activity of the methionine amide analogue of enkephalin¹² over Met-enk. In tissues where peptidyl dipeptidase is concentrated, enkephalins may be readily degraded by this enzyme. Furthermore, by reacting with the enzyme enkephalins may interfere with the metabolism of other active peptides. Hypothetically they may inhibit the release of angiotensin II from angiotensin I or the inactivation of kinins by competing for the same enzyme.

Cultured human endothelial cells, however, inactivate enkephalins primarily by the action of an aminopeptidase, as demonstrated by the release of Tyr¹. The degradation of enkephalins by a similar aminopeptidase has been shown by using purified enzyme,¹³ plasma, brain homogenates^{12,14}, or a membrane fraction of rat brain² as sources of enzyme.

Our experiments indicate that rapid inactivation of enkephalins may be due to at least two different enzymes present in tissues. These enzymes presumably are localized in different structures of the cells. A peptidyl dipeptidase which is a component of plasma membrane of various cells⁶ may degrade enkephalins by liberating a C-terminal dipeptide and an aminopeptidase in endothelial cells may cleave peptides that they are taken up as blood flows past the endothelial surface at the N-terminal end,

Legends

Figure 1. Hydrolysis of Leu⁵- and Met⁵-enkephalin by purified hog kidney peptidyl dipeptidase (angiotensin I converting enzyme or kininase II) to Tyr-Gly-Gly and Phe-Leu or Phe-Met. The specific inhibitor SQ 14225 (10^{-6} M) inhibited the reaction. Peptides were incubated with the enzyme up to 60 min in presence and absence of inhibitor. a. Silica gel t.l.c. plate. Solvent: sec-butanol: acetic acid: water (4:1:5). Separation time 3 h. b. Aluminum oxide t.l.c. plate. Solvent: chloroform: methanol: NH_4OH (20:20:9). Separation time 1.5 h. The plates were sprayed with ninhydrin.

Figure 2. Hydrolysis of Met⁵-enkephalin by purified peptidyl dipeptidase determined in an automatic amino acid analyzer in absence (0————0) and in presence (0-----0) of the specific inhibitor SQ 20881 (10^{-4} M).

Figure 3. Cleavage of Tyr¹ from Leu⁵-enkephalin by human vascular endothelial cells (a) and by purified leucine aminopeptidase (b). Inhibitor: o-phenanthroline (10^{-3} M). Silica gel plates, conditions as in Fig. 1.

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